



## Training Course specification of Parasitology

### 1-Basic information

<b>Course title :</b>	Training Course specification of Parasitology
<b>Academic year:</b>	3rd academic year (2014/2015)
<b>Programme title:</b>	Bachelor of Veterinary Medical Sciences
<b>Contact hours /week/semester:</b>	See training program specification

### 2-Professional information

#### 1- Overall aims of course

The training program aimed to produce trained veterinary students:  
Able to deal with the broad spectrum of specimens and problems encountered in general veterinary parasitology.  
Acquire skills and competence in a particular species sufficient to lead a diagnostic team in this area if required.

#### 2- Intended learning outcomes of course (ILOs)

##### a-Knowledge and understanding

**By the end of this course the student should be able to:**

- 1) Develop of the clinical and molecular parasitology and of slides preparations to be able to interact appropriately with colleagues responsible for those aspects of technical work
- 2) Acquire knowledge of parasitology including sampling, signs on animals and rapid methods of diagnosis.
- 3) Acquire the habit of lifelong learning by a combination of reading, literature searches, consultation with colleagues, attendance at scientific meetings, and the presentations.

##### b-Intellectual skills

**By the end of this course the student should be able to :**

- 1) Interpret both clinical signs on animals, macroscopic and microscopic from the findings of post-mortem examinations.

### c-Professional and practical skills

**By the end of this course the student should be able to:**

- 1) Acquire Understanding of information technology sufficient to be able to use computers for producing parasitological reports, to search databases, access e-mail and internet services

### d-General and transferable skill

**By the end of studying the course, the student should be able to**

- 1) Deal with animal owner and understand the field language to take appropriate case history.
- 2) Acquire Management and communication skills in order to interact appropriately with medical, scientific and technical colleagues in the workplace and eventually to function as a team leader, if so requested.

### **3-Topics and contents**

<b>Topic</b>		
<b>I- Necropsies Training</b> <ul style="list-style-type: none"><li>• Training in the performance and reporting of necropsies.</li><li>• Supervised performance and reporting of necropsies including basic examination of the systems, parasitological sampling, and evaluation of available clinical data.</li></ul>		
<b>II- Diagnostic parasitological Training</b> <ul style="list-style-type: none"><li>• Training in the performance of the parasitological techniques</li><li>• Training in diagnostic reading of prepared microscopic slide.</li><li>• Training in the principles of parasites mounting.</li><li>• Practice in writing parasitological reports including advice on their content, composition, diagnostic comments and suggested treatment for the case.</li></ul>		
<b>III Detailed description of the training program:</b>		
<b>First week</b>		
<b>Day</b>	<b>First session</b>	<b>Second session</b>
1 <sup>st</sup> day	Sampling	Case study (parasitology)
2 <sup>nd</sup> day	Sedimentation and Floatation method	Fecal analysis (helminth and protozoa)
3 <sup>rd</sup> day	Larval fecal culture	McMaster and Baermann Tech.
4 <sup>th</sup> day	Blood and other body fluids examination	Detection of protozoa and filarial worms
5 <sup>th</sup> day	Skin scraping and tissue exam.	Mange diagnosis
6 <sup>th</sup> day	Necropsy examination and reporting of small animals	Case study (Necropsy)
<b>Second week</b>		
<b>Day</b>	<b>First session</b>	<b>Second session</b>
1 <sup>st</sup> day	Sampling	Case study (parasitology)

2 <sup>nd</sup> day	Sedimentation and Flootation method	Fecal analysis (helminth and protozoa)
3 <sup>rd</sup> day	Larval fecal culture	McMaster and Baermann Tech.
4 <sup>th</sup> day	Blood and other body fluids examination	Detection of protozoa and filarial worms
5 <sup>th</sup> day	Skin scraping and tissue exam.	Mange diagnosis
6 <sup>th</sup> day	Necropsy examination and reporting of birds	Case study (Necropsy)

### Third week

Day	First session	Second session
1 <sup>st</sup> day	Sampling	Case study (parasitology)
2 <sup>nd</sup> day	Sedimentation and Flootation method	Fecal analysis (helminth and protozoa)
3 <sup>rd</sup> day	Larval fecal culture	McMaster and Baermann Tech.
4 <sup>th</sup> day	Blood and other body fluids examination	Detection of protozoa and filarial worms
5 <sup>th</sup> day	Skin scraping and tissue exam.	Mange diagnosis
6 <sup>th</sup> day	Necropsy examination and reporting of fish	Case study (Necropsy)

### Forth week

Day	First session	Second session
1 <sup>st</sup> day	Sampling	Case study (parasitology)
2 <sup>nd</sup> day	Sedimentation and Flootation method	Fecal analysis (helminth and protozoa)
3 <sup>rd</sup> day	Larval fecal culture	McMaster and Baermann Tech.
4 <sup>th</sup> day	Blood and other body fluids examination	Detection of protozoa and filarial worms
5 <sup>th</sup> day	Skin scraping and tissue exam.	Mange diagnosis
6 <sup>th</sup> day	Methods of parasites Mounting	Collected samples from necropsies

## 4-Teaching and learning methods -

- 4.1 Sessions of sampling and necropsy training.
- 4.2. Sessions of laboratory diagnostic parasitological techniques for Identification of different parasites.

## 6-Student assessment

See training program specification

Program coordinator	Head of department
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